06.11.20

### **CURRICULUM VITAE**

### **GENERAL**

### Prof. Teuta Pilizota

School of Biological Sciences, College of Science and Engineering, University of Edinburgh

### CAREER:

2020-	Professor of biophysics, School of Biological Sciences, University of Edinburgh
2019-	Director of Research, School of Biological Sciences, University of Edinburgh
2018-2020	Reader in biophysics, School of Biological Sciences, University of Edinburgh
2013-2018	Chancellor's Fellow with tenure at School of Biology, University of Edinburgh

2008-2012 Postdoctoral research fellow at Princeton University. Working with Prof Joshua W Shaevitz
 2007-2008 Postdoctoral research fellow at the University of Oxford. Working together with Dr. Richard

M Berry and Prof. Judith P Armitage

In April-July 2011 I had a short-term disability leave due to Staphylococcus Aureus hip bone infection and bacteraemia

# UNIVERSITY EDUCATION:

**2002 – 2007** D.Phil. in Biological Physics, University of Oxford, Department of Physics.

Under supervision of Dr. Richard M. Berry

1997 - 2002 Diploma in Physics, University of Zagreb, Faculty of Science, Department of

Physics. grade average 4.82 (on a scale from 1 to 5)

## **TEACHING**

**2016, 2017** Lecturer at Hands-on Research in Complex Systems School, International Centre for

Theoretical Physics, Trieste, teaching included practical experiment and career development training sessions and lectures for researchers from developing countries

2016-2017 Lecturer, Microbial World II (2<sup>nd</sup> year), School of Biology, University of Edinburgh, UK
2014- Lecturer, Molecular Microbiology III (3<sup>rd</sup> year), School of Biology, University of Edinburgh,

UK

2013- Lecturer, Novel Approaches in Biotechnology and Membrane Biology (4th year), School of

Biology, University of Edinburgh, UK

Tutor, preparing adult learners (age 25-40) for GED exams, Princeton, New Jersey, USA.

2009 – 2010 Lecturer, Math 135, part of Mercer County Community College and The College of New

Jersey AA degree, Edna Mahan Correctional Center for Women in Clinton, New Jersey,

USA.

2006 – 2008 Marker, Biophysics course, Department of Physics, University of Oxford

2004 – 2005 Demonstrator, Biological physics practical course, Department of Physics, University of

Oxford

# TAUGHT PROGRAMS STUDENT SUPERVISION (as an independent PI):

Master Students: Supervised two ERASMUS exchanged student from University of Zagreb,

Department of Biotechnology in 2014 and Department of Electrical Engineering and Computing in 2018, 3 School of Biological Sciences (Biotechnology and Synthetic Biology Master program) and 2 School of Physics and Astronomy Master students at the University of Edinburgh 2016-2019, one Master by Research student at School of Biological Sciences, University of Edinburgh and a MEng student from University of

Glasgow, Bioengineering program in 2019

Under Graduate Students: Supervised 10 honors students from the University of Edinburgh in 2014-

2020 (8 in Biotechnology honors and 2 in Biochemistry)

Supervised two summer project students from Department of Life

Sciences, Peking University, Bejing, summer 2014

## RESEARCH

EXTERNAL RESEARCH FUNDING:

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2020	IBioIC Feasibility Fund (with OGI bio Ltd), co-I, £33,6k				
2020-2021(22	) ONR Global X Challenge: Bacterial Flagellar Motor as a multimodal biosensing chip, PI				
	\$252k (+\$288k)				
2020-2024	IBioIC: Technology development for in situ imaging of microbial cultures, with OGI Bio Ltd				
	PI, £136k				
2020 -2023	A Physiological Approach To Understanding Osmotically Induced Growth				
2020 2020	Modulation: Leverhulme Trust, <b>PI, £177k</b>				
2019	BBSRC Pathfinder and iCURE, <b>PI, £70k</b>				
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2018-2021	Office of Naval Research (ONR) and Defense Advanced Research Projects Agency (DARPA), Bacterial flagellar motor as fast synthetic biosensor, <b>PI, \$390.6k</b>				
2015-2016	Royal Society Brian Mercer Award, Microfluidic platform for monitoring product accumulation				
	and 'health state' of bacterial hosts cells during bioproduction, PI, £30k				
2015-2018	HFSP: Revealing bacterial free energy dynamics during loss of viability, PI, \$1.05M				
2015-2020	BBSRC/EPSRC/MRC: Synth Mammalian: Edinburgh Mammalian Synthetic Biology				
	Research Centre, <b>co-I, £13.2M</b>				
2015-2019					
2015-2019	BBSRC iCASE: Engineering bacterial hosts cells for robust growth at high external				
	osmolarities, with INEOS as an industrial partner, PI, £91k				
2015-2018	Cunningham Trust: Measuring phenotypical strategies bacteria employ to sustain viability				
	under antibiotic treatment and identifying optimal strategies needed to combat them, PI,				
	£71k				
2015-2018	IBioIC: Preventing unwanted cytoplasmic leakage in downstream processing, with				
	FujiFilm, PI, £71k				
2014-2016	Crossing Biological Membrane BBSRC Network in Industrial Biotechnology and Bioenergy				
	Proof of Concept Award and Business Interaction Voucher: Using <i>E. coli</i> turgor pressure				
	regulation to optimize product excretion and prevent unwanted cytoplasmic leakage, with				
	FujiFilm Diosynth Biotechnologies as industrial partner, <b>PI, 30k</b>				
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# INTERNAL RESEARCH FUNDING AND SMALLER TRAVEL GRANTS:

2019-2020	Technology development for direct imaging of microbial cells in a
	Microbioreactor, EPSRC Impact Acceleration Award, PI, £55k
2019-2020	Fully automated, aerated and affordable microbioreactor technology, BBSRC Impact
	Acceleration Award, PI, £19.5k
2018	SUPA Distinguished Visitor Award, £1.7k
2018	Improving desert agriculture through insights in Nitrogen fixing bacteria, BBSRC GCRF
	Impact Acceleration Award, PI, £2.7k
2017	Information processing by bacterial chemotactic network, EPSRC NetworkPlus
	Emergence and Physics Far From Equilibrium, PI, 1.9k
2015	Microfluidic platform for monitoring product accumulation and 'health state' of bacterial
	hosts cells during bioproduction, EPSRC Impact Acceleration Award, PI, £15.8k
2015	Assessing growth of individual lactic acid bacterial cells in beer, EPSRC Impact
	Acceleration Award, PI, £4.5k
2015	Capturing dynamic cellular responses to mechanical stimuli, ISSF2 University of
	Edinburgh fund, co-I, £29.5k
2013	International Exchange Program, Taiwan MOS Bilateral Award, Royal Society of
	Edinburgh PI, £2.7k
2013	Novel approaches to bacterial stress response, ISSF University of Edinburgh fund, PI,
	£20k

# RESEARCH STUDENT SUPERVISION:

Currently supervising 5 Ph.D. students (one in his 4<sup>th</sup> year, and three in 2<sup>nd</sup> year and one just started his 1<sup>st</sup> year).
Seven Ph.D. student successfully defended their thesis since 2013 (4 in

2019)

# **KNOWLEDGE EXCHANGE AND IMPACT**

FUNDING RECEIVED FOR TRANSLATIONAL WORK WITH INDUSTRY:

2020 IBioIC Feasibility Fund (with OGI bio Ltd), co-I, £33,6k

2020-2024	IBioIC: Technology d	levelopment for <i>in situ</i> imaging	of microbial cultures	with OGI Bio Ltd
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PI, £136k

2015

**2019** EPSRC DTP CASE conversion scholarship with Chr. Hansen

2015-2016 Royal Society Brian Mercer Award, Microfluidic platform for monitoring product

accumulation and 'health state' of bacterial hosts cells during bioproduct, PI, £30k

2015 Microfluidic platform for monitoring product accumulation and 'health state' of bacterial

hosts cells during bioproduction, EPSRC Impact Acceleration Award, PI, £15.8k

Assessing growth of individual lactic acid bacterial cells in beer, EPSRC Impact

Acceleration Award, PI, £4.5k

2015-2019 BBSRC iCASE: Engineering bacterial hosts cells for robust growth at high external

osmolarities, with INEOS as an industrial partner, PI, £91k

2015-2018 IBioIC: Preventing unwanted cytoplasmic leakage in downstream processing, with

FujiFilm, PI, £71k

2014-2016 Crossing Biological Membrane BBSRC Network in Industrial Biotechnology and Bioenergy

Proof of Concept Award and Business Interaction Voucher: Using *E. coli* turgor pressure regulation to optimize product excretion and prevent unwanted cytoplasmic leakage, with

FujiFilm Diosynth Biotechnologies as industrial partner, PI, 30k

### SPIN-OUT COMPANIES AND PATENTS:

**2020** ŌGI Bio Ltd (https://www.ogibio.com/) startup registered under company house

2019 BBSRC Pathfinder and iCURE funding for market research, PI, £65k
2020-2021 Innovate UK: ŌGI Bio Ltd iCURE follow on fund application, co-I, £249k

#### DISSEMINATION OF GOOD PRACTICE IN KNOWLEDGE EXCHANGE:

2016 Collaboration with WEST brewery featured in Infinite Magazine, Edinburgh Research and

Innovation (http://www.ed.ac.uk/files/atoms/files/infinite-magazine-2016.pdf)

2016 Collaboration with FujiFilm DioSynth featured in BBSRC CBMNet (Crossing Biological

Membranes Network) documentary https://www.youtube.com/watch?v=RtbNV73FGjs

# SCIENCE OUTREACH ACTIVITIES:

**2016** Featured in the BBC Arabic Science documentary

(https://www.youtube.com/watch?v=qs6cOxgBJ9U&list=PL63lwGZ 8vsmtS9CyVZSKiWL

QmGw7H4IO&index=1)

2016 Feature in the leading daily Croatian national newspaper, discussing the profession of a

modern academic (http://www.jutarnji.hr/life/znanost/biti-znanstvenik-danas-covjek-koji-je-otkrio-bozju-cesticu-tvrdi-danas-ne-bih-mogao-dobiti-posao-u-akademskoj-zajednici.-

je-li-u-pravu/3709260/)

2013 Contributing to the School of Biology showcase at the Edinburgh International Science

Festival, National Museum of Scotland, Edinburgh, UK

2013 Invited speaker at the Summer School of Science aimed at high-school students,

Pozega, Croatia

2012 Team leader, Microscopy exploration station for middle school students, Science and

Engineering EXPO, Princeton University, USA

2004 – 2007 Connect project leader, Connect is a project of NGO znanost.org (science.org) whose

goals and activities include network building for knowledge professionals, reversing the brain-drain into the skills-gain, organizing annual gatherings of Croatian scientists working in the country and abroad, funding student projects and research experience

**2003 – 2007** Executive board member, NGO znanost.org (science.org)

2002 - 2007 NGO znanost.org (science.org), member, znanost.org is a non-governmental

organization whose function is to promote education, science and knowledge-based

values in Croatian society through a hands-on approach

# ACADEMIC LEADERSHIP, MANAGEMENT, AND CITIZENSHIP

2020 Co-director of Hands-on Research in Complex Systems School, August, International

Centre for Theoretical Physics, Trieste, Italy (cancelled)

**2019-2021** Organizing committee: 3<sup>rd</sup> Electrical Cell Biology Workshop

2019- Research Director, School of Biological Sciences2019 BBSRC Committee B Pool of experts member

2017-2018 Organizing committee member of Physics of Cells: From Biochemical to Mechanical
 2017-2018 European Federation of Biotechnology meeting on Microbial stress, April 2018,

International Scientific Advisory Board Member

2017 Global Challenges Research workshop with Mohammed V University in Rabat, Morocco,

October 2017, organizer

2017-2018 School of Biological Sciences, UoE Internationalization Committee Member

2016-2017 Microbiology Society Annual Conference, April 2017, Synthetic and Systems Biology

Session organizer

**2016-** BBSRC Committee D Pool of experts member

2014-2018 School of Biological Sciences, UoE Research Committee Member

2014 External Master Thesis Examiner, University of Zagreb, Department of Biotechnology
 2013- Peer-reviewer for grant proposals to Swiss National Science Foundation, BBSRC, Netherlands Organization for Scientific Research, Cunningham Trust and Carnegie Trust
 2013- Peer-reviewer for several international, interdisciplinary journals: PNAS, Science

Advances, Scientific Reports, Applied Physics, Biological Physics, Frontiers, etc.

# **EXTERNAL RECOGNITION/ESTEEM**

2019 External Examiner, University of Oxford, Department of Zoology
 2019 External Examiner, University of York, Department of Biology
 2018-2020 Visiting Researcher, Department of Physics, University of Oxford

2018 Chief of Naval Research USA, Rear Admiral David A. Hahn visit to Pilizota lab
 2007 Merit Award for exceptional performance, Department of Physics, University of Oxford
 2007 Daiwa Adrian Prize for UK-Japan joint collaboration in *Analysis of the Mechanism and*

Structural Dynamics of the Bacterial Flagellar motor, UK scientific research team

# KEYNOTE AND INVITED SPEAKER AT INTERNATIONAL MEETINGS:

**2021** 87th Harden Conference: Single-molecule bacteriology II, Oxford, UK (Invited Speaker)

2020 'Microbial Cell Biology' meeting, October, Berlin, Germany, (Invited Speaker, postponed)

2020 UK/USA Synthetic Biology Showcase, May, UK (Invited Speaker, cancelled)

2020 Naoninbio conference, April, Guadeloupe, France, (Invited Speaker, postponed)

2020 'Flagellar meeting', March Taiwan, (Invited Speake, Cancelled)

**2020** EuroMicropH COST action open meeting (Invited Speaker)

2020 'New Physical Models for Cell Growth', January, Aspen Centre for Physics, USA (Invited Speaker)

**2019** SynBioUK, December, UK (Invited Speaker)

2019 'Biophysics of Infection and Immunity: From Molecules to Cells to Tissues' symposium, University of York, November, UK (Invited Speaker)

**2019** UK Biofluids Special Interest Group Meeting, September, University of Warwick (Invited Speaker)

2019 13th Annual q-Bio Conference, August, San Francisco, USA (Invited Speaker)

**2019** HFSP Annual Meeting, July, Tsukuba, Japan (Invited Speaker)

2019 UK/USA Synthetic Biology Showcase, Williams Formula 1 Conference Centre, Wantage, May UK (Invited Speaker)

2019 The physics of microorganisms II, April, Institute of Physics, London, UK (Invited Speaker)

2019 2<sup>nd</sup> Electrical Cell Biology Workshop, Warwick University, UK (Invited Speaker)

2019 Quantitative Bacterial Cell Biology Symposium, January, Institute Pasteur, Paris (Invited Speaker)

2018 Biochemical Society Harden Conference on Single-Molecule Bacteriology, August, Oxford, UK (Invited Speaker)

2018 Integrative Cell Models for Disease Intervention, Banff International Research Station for Mathematical Innovation and Discovery, June Canada, (Invited Speaker)

2018 European Federation of Biotechnology meeting on Microbial stress, April, Ireland (Invited Speaker)

2018 American Physical Society March Meeting, L.A., USA (Invited Speaker)

2017 Nanofluidics in Biological Systems Workshop, Durham University, September (Invited Speaker)

2017 19th IUPAB and 11th European Biophysical Society Meeting Annual Conference, July, HFSP session (Invited Speaker)

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- **2017** Membrane Engineering Of Lipids And Proteins For Industrial Biotechnology And Bioenergy, June, Glasgow (Invited Speaker)
- 2016 Physics Meets Biology, September 2016, Clare College, Cambridge, UK (Invited Speaker)
- **2016** Knaw Biophysics Meeting, Amsterdam (Invited Speaker)
- 2015 BioSynSys2015 conference, University Paris Diderot, Paris, France (Keynote Speaker)
- **2015** Integrative Cell Models: Bridging Microbial Physiology and Systems Biology, Lorentz Centre, Leiden University, Leiden, Netherlands (Invited Speaker)
- 2013 Croatian Physical Society Meeting, Primosten, Croatia (Invited Speaker)

### INVITED SPEAKER AT SUMMER SCHOOLS:

- 2020 MicroQUANT, Institut d'études Scientifiques de Cargèse, June, France
- 2020 Hands on Research in Complex Systems School, Aug-Sep 2020 ICTP, Trieste, Italy (lecturer)
- 2017 CM-CDT and Higgs Summer school, University of St. Andrews, August, St. Andrews UK
- **2017** Hands-on Research in Complex Systems School, August, International Centre for Theoretical Physics, Trieste, Italy (Invited Lecturer)
- **2016** From Molecules to Systems 2016 Winter School, University of Oxford, St. Catherine's College, Oxford, UK (Invited Speaker)
- 2016 Hands-on Research in Complex Systems School, July, International Centre for Theoretical Physics, Trieste, Italy (Invited Lecturer)

### CONTRIBUTING SPEAKER AT INTERNATIONAL MEETINGS:

- 2014 Stochastic Biology Conference: from Cells to Populations, Institute of Science and Technology, (IST) Austria
- 2014 16th European Congress on Biotechnology, Edinburgh, UK
- 2010 "Microbial Stress Response" Gordon Research Conference, Mount Holyoke College, MA, USA
- 2007 51st Annual Meeting of American Biophysical Society, Baltimore USA
- 2005 Bionanotechnology Collaboration Conference, Tokyo Japan, December 2006
- 2005 Croatian Biophysical Society, Zagreb, Croatia
- 2005 15th Congress of the IUPAB-5th EBSA Congress, Montpellier France

# INVITED SPEAKER AT UK AND INTERNATIONAL UNIVERSITIES:

- 2017 University College London, Department of Physics, UK
- 2016 Newcastle University, Institute for Cell and Molecular Biosciences
- 2016 University of Durham, Department of Physics, UK
- 2016 Leeds University, Department of Physics, Leeds, UK
- 2016 Warwick University, Centre for Synthetic Biology, Warwick, UK
- 2016 Bristol University, Department of Applied Mathematics, Bristol, UK
- 2016 Imperial College London, Department of Structural Biology, UK
- 2016 National University of Singapore, Centre for Synthetic Biology, Singapore
- 2016 National University of Singapore, Department of Physics, Singapore
- 2015 MRC Clinical Sciences Centre, Imperial College London, London, UK
- 2015 Department of Biochemistry, University of Oxford, Oxford, UK
- 2014 Department of Physics, University of Zagreb, Zagreb, Croatia
- 2014 College of Life Sciences, University of Dundee
- 2014 Institute of Microbiology and Infection, School of Biosciences, University of Birmingham, Birmingham, UK
- 2014 Department of Physics, National Central University, Jhongli, Taiwan
- 2013 Department of Physics, University of Sheffield, Sheffield, UK
- 2012 National Institute for Medical Research, London, UK
- 2012 Department of Biology, York University, York, UK
- 2012 School of Biology, University of Edinburgh, Edinburgh, UK
- 2010 Department of Physiology and Biophysics, University of Washington, Seattle, USA
- 2010 Clarendon Laboratory, Department of Physics, University of Oxford, Oxford UK
- 2008 Chemical and Process Engineering Department, Sheffield University seminar series, Sheffield UK
- 2007 Institute of Scientific and Industrial Research, Department of Single Molecule Biophysics, Osaka University, Osaka, Japan

# **LIST OF PUBLICATIONS:**

Schwarz-Linek J, Krasnopeeva E, Douarchec C, Arlt J, **Pilizota T**, Poon WCK and Martinez VA 'Motility turns sour: how a combination of environmental factors prevents swimming in dense suspensions of E. coli' (In preparation)

Wong F\*\*, Wilson S, Helbig R, Hegde S, Aftenieva O, Zheng H, Liu C, **Pilizota T**, Garner EC, Amir A\*\*, Renner LD\*\* 'Single-cell lytic probing of the bacterial cell envelope' (Submitted)

Teraddot G, Krasnopeeva E, Swain S, **Pilizota T**. 'Escherichia coli's ability to maintain intracellular pH changes with the Proton Motive Force' (In preparation)

Voliotis M, Rosko J, **Pilizota T**, Liverpool T. 'Steady state running rate sets the speed and accuracy of accumulation of swimming bacterial populations' (https://arxiv.org/abs/2007.08335, Under Review)

Mancini L, **Pilizota T**\*\*. 'Environmental conditions define the energetics of bacterial dormancy and its antibiotic susceptibility' (https://www.biorxiv.org/content/10.1101/2020.06.18.160226v1, Under Reivew)

Krasnopeeva E, Barboza-Perez U E, Rosko J, **Pilizota T\*\***, Lo C J\*\* 'Bacterial Flagellar Motor as a Multimodal Biosensor' **Methods** 2020; In press (https://doi.org/10.1016/j.ymeth.2020.06.012)

Schofiel Z\*, Meloni G\*, Tran P, Zerfass C, Sena G, Hayashi Y, Grant M, Contera SA, Minteer SD, Kim M, Prindle A, Rocha P, Djamgoz MBA, **Pilizota T**, Unwin PR, Asally M\*\*, Soyer OS\*\* 'Bioelectrical understanding and engineering of cell biology' *J. R. Soc. Interface*, 2020;17: 20200013

Paraschiv A, Hegde S, Ganti R, **Pilizota T**, Saric A\*\* Dynamic clustering regulates activity of mechanosensitive membrane channels, **Physical Review Letters**, 2020; 124:048102

Mancini L, Tian T\*, Terradot G\*, Pu Y, Li Y, Lo CJ, Bai F, **Pilizota T\*\*** A general work-flow for characterization of Nernstian dyes and their effects on bacterial physiology, **Biophys J**, 2020;118(1): 4-14

Wang YK\*, Krasnopeeva E\*, Bai F, **Pilizota T\*\***, Lo CJ\*\* Comparison of Escherichia coli surface attachment methods for single-cell, in vivo microscopy, **Scientific Reports**, 2019;9:19418

Arlt J, Martinez VA, Dawson A, **Pilizota T**, Poon WCK\*\* 'Dynamics-dependent density distribution in active suspensionsons' *Nature Communications* 2019;10: 2321

\*Krasnopeeva E, Lo CJ, **Pilizota T**\*\* 'Single-cell bacterial electrophysiology reveals mechanisms of stress induced damage' **Biophys. J.** 2019;116(12): 2390-2399

**Pilizota T** and Ya-Tang Yang\*\*. Flexible and affordable microbial cultivation techniques for synthetic and systems biology, *Frontiers in Microbiology*, 2018;9:1666 doi=10.3389/fmicb.2018.01666

\*Arlt J, Martinez V A, Dawson A, **Pilizota T** and Poon WCK\*\*. Spatially-controlled activity of light-driven bacteria. **Nature Communications** 2018:9: 768

Rosko J, Martinez V A, Poon WCK and **Pilizota T\*\***. Osmotaxis in *Escherichia coli* through changes in motor speed *PNAS* September 2017; doi: 10.1073/pnas.1620945114

Stevenson K, McVey A F, Clark I B N, Swain P S and **Pilizota T\*\***. General calibration of microbial growth in microplate readers. **Scientific Reports.** 2016; 6:38828

Swain P S\*\*, Stevenson K, Leary A, Montano-Gutierrez L F, Clark I B N, Vogel J and **Pilizota T**. Inferring time-derivatives, including cell growth rates, using Gaussian processes. *Nature Communications*. 2016; 7:13766

\*Buda R\*, Liu Y\*, Yang J\*, Hegde S\*, Stevenson K, Bai F\*\* and **Pilizota T\*\***. Dynamics of *Escherichia coli's* passive response to sudden decreases in external osmolarity. *PNAS*. September 2016, doi:10.1073/pnas.1522185113

Fletcher KA, **Pilizota T**, Rhys-Davies P, French, CE. Characterization of the effects of n-butanol on the cell envelope of *E. coli.* **Applied Microbiology and Biotechnology.** September 2016, 1-7. doi:10.1007/s00253-016-7771-6

Schwarz-Linek J, Arlt J, Jepson A, Dawson A, Vissers T, Miroli D, **Pilizota T**, Martinez VA and Poon W. *Escherichia coli* as a model active colloid. *Colloids and Surfaces B: Bionterfaces.* 2016. 137:2-16

**Pilizota T\*\***, Shaevitz JW. Origins of cell shape and growth rate changes at high external osmolarity. **BIOPHYS J** October 2014, 107(8):1962-1969

**Pilizota T,** Shaevitz JW. Plasmolysis and cell shape depend on solute outer membrane permeability during hyperosmotic shock in *Escherichia coli*. **BIOPHYS J** 18 June 2013, 104(12):2733-2742

Lo CJ, Sowa Y, **Pilizota T**, Berry RM. The mechanism and kinetics of a sodium-driven bacterial flagellar motor. **PNAS**. July 2013.110(28):E2544-51

Bilyard T\*, Nakanishi-Matsui\* M, Steel B, **Pilizota T**, Nord A, Hosokawa H, Futai M, Berry RM. High-resolution single-molecule characterization of the enzymatic states in *Escherichia coli* F<sub>1</sub>-ATPase. *PHIL TRANS R SOC. B* 24 December 2012. 368(1611):20120023.

**Pilizota T**, Shaevitz JW. Fast, multiphase volume adaptation to hyperosmotic shock by *Escherichia Coli.* **PLoS ONE** 2012 Apr; 7(4): e35205

\*Fan B\*, Branch RW\*, Nicolau DV\*, **Pilizota T**, Maini PK, Berry RM. Conformational spread as a mechanism for cooperativity in the bacterial flagellar switch. *SCIENCE*. 2010 Feb; 327(5966): 685-9. (Mentioned in Perspectives: An Ensemble View of Allostery, Vincent J. Hilser, *Science* 5 February 2010: 653-654). \*These authors contributed equally

\*Pilizota T\*, Brown M\*, Leake MC, Branch RW, Berry RM, Armitage JP. A molecular brake, not a clutch, stops the *Rhodobacter sphaeroides* flagellar motor. *PNAS*. 2009 Jul;106(28):11582-7. (Mentioned in 'In This Issue', PNAS 2009 106 (28) 11427-11428) \* These authors contributed equally

Inoue Y, Lo CJ, Fukuoka H, Takahashi H, Sowa Y, **Pilizota T**, Wadhams G, Homma M, Berry RM, Ishijima A. Torque-speed relationships of Na<sup>+</sup>-driven chimeric flagellar motors in *Escherichia coli. J MOL BIOL.* 2008 Mar; 376(5):1251-9.

Lo CJ, Leake MC, **Pilizota T**, Berry RM. Non-equivalence of membrane voltage and ion-gradient as driving forces for the bacterial flagellar motor at low load. **BIOPHYS J.** 2007 Jul; 93(1):294-302.

**Pilizota T**, Bilyard T, Bai F, Hosokawa H, Futai M, Berry RM. A programmable optical angle clamp for rotary molecular motors, *BIOPHYS J.* 2007 Jul; 93(1):264-275

**Pilizota T**, Lucic B, Trinajstic N. Use of variable selection in modeling the secondary structural content of proteins from their composition of amino acid residues. **J CHEM INF COMPUT SCI.** 2004 Jan-Feb;44(1):113-21.

# **Book contributions:**

**Pilizota T**, Sowa Y, Berry RM. Chapter: Single-Molecule Studies of Rotary Proteins in Handbook of Single-Molecule Biophysics, Springer. 2009. Editors: Peter Hinterdorfer and Antoine van Oijen